Rational design of a material for rapid colorimetric Fe²⁺ detection

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ABSTRACT: We report on the rational design of a novel TiO₂-based screen printed material suitable for sensitive and selective detection of iron ions in water. This includes the synthesis and characterization of large mesoporous TiO₂ nanostructures, screen-printing of titania thick films on glass surfaces and their functionalization with 2,2':6',2"-terpyridin-4'-ylphosphonic acid (terpy). The ultra-high affinity between iron ions and TiO₂-anchored terpy receptor makes this system potentially applicable for analysis of the iron content in environmental, food, biological, and biomedical systems by a readily quantifiable colour change. Rapid (<30 s) colour change of the material from white to magenta permits easy detection of as low as 0.3 ppm of Fe²⁺ by the naked eye. Colour change intensity depends on the nature of the nanoparticles, the overall TiO₂ film thickness, and the Fe²⁺ concentration. The material was characterized using profilometry, diffuse reflectance UV-Vis spectroscopy, and X-ray photoelectron spectroscopy (XPS) before and after treatment with aqueous solutions of Fe²⁺. The designed material shows colour reversibility upon treatment with EDTA solutions, which allows for multiple re-uses of the same film with no effect on sensitivity.

KEYWORDS: TiO₂, screen printed films, terpy, iron detection.

HIGHLIGHTS:

Depositing of terpy-based ligand, L, on TiO₂ support results in hybrid material able to selectively react with Fe²⁺.

Impregnation of iron (II) into material leads to distinct colour change from white to magenta.

A novel material based on screen-printed mesoporous TiO_2 beads functionalized by L was designed and characterized.

The material is able to quantify 0.3 to 5 ppm of Fe^{2+} in aqueous solutions.

1. Introduction

Inexpensive fast detection and quantification of metal cations in aqueous solutions is in high demand for a variety of environmental, industrial, forensic, biological and health applications. Especially interesting are systems that allow colorimetric detection to be performed by non-trained personal in a field condition by the naked eye [1, 2]. Thus, the development of materials for such purpose is crucial [3]. Incorporation of sensing receptors on solid supports is a very promising strategy for the development of this type of material. The most common approach is the integration of the functional motif of interest, i.e. a sensing receptor into a polymeric matrix or formation of

self-assembled monolayers (SAMs) containing a sensing receptor on the surface by utilization of an appropriate linker. Various derivatives of 2,2':6',2"-terpyridine (terpy) have been developed and implemented as such sensing receptors as summarized in seminal works by Constable [4] and Hancock [5]. Terpy-based molecules were utilized to identify various metal cations [5-7] and cation-anions pairs [8, 9]. To adjust polarity and therefore solubility, as well as to enhance sensing characteristics, terpy motifs have been combined into a variety of organic, organometallic and supramolecular architectures. These structures were applied for metal ion sensing [10-12], counter anion sensing [8, 13, 14], sensing of bio-molecules [15, 16], and for bio-imaging [17]. Incorporation of terpy derivatives into polymeric matrices often results in the formation of functional polymeric materials that can be employed as effective sensors [18]. For example, solutions of polyiminofluorene based polymer decorated with terpy groups were utilized as a highly sensitive and specific fluorimetric zinc sensor in organic solvent media [10]. Very recently, Garcia et al. reported the incorporation of terpy motif into water soluble acrylic polymers for colorimetric detection of Fe³⁺, Co³⁺, Sn²⁺, and Cu²⁺ in solution and on solid film [19]. In addition, they reported on the design of polymeric materials able to detect the sub-micromolar amounts of Fe³⁺ and Co³⁺ and micromolar amounts of Cu²⁺ and Sn²⁺. Remarkably, varying the amount of terpy incorporated into the polymer allows for the effective tuning of the selectivity of the resulting polymeric material towards ion sensing. Moreover, a terpy-functionalized poly(ionic liquid) has been utilized for coordination and selective recognition of europium ions that further can be employed in the preparation of novel luminescent materials [20, 21].

Besides the successful integration of terpy into soft materials and polymers, incorporation of this functional component into solid support is an important step for the creation of materials for molecular electronics and photonics [22], electrochromic materials [23], and solar cells. Terpy-motifs have been anchored onto a large number of solid supports resulting in functional materials with enhanced stability and excellent light absorption properties [18, 23-29]. Nishihara et al. have achieved long range electron transport for molecular electric wires by assembling bis(terpyridine) metal complexes into molecular wires on gold surfaces. Successively, they have also used the same molecular motif to create electrochromic responsive nanostructures on the water-organic interface [30, 31]. Terpy–based macroligands were used to prepare yellow-green emitting Zn²⁺ metallo-polymer complexes with advanced photophysical properties for OLED applications [32]. Finally, the recent work of Li and Han shows that immobilization of non-planar terpy motifs on graphene-based porous materials through an azide-alkyne "click" reaction enhances porosity and carbon dioxide gas sorption capacity [33].

It is well documented that the terpy-core motif allows visual differentiation of iron cations due to a rapid color change from colorless to deep magenta and this effect has been widely employed for the determination of Fe^{2+} . Remarkably, after more than 80 years since the discovery of the first Fe^{2+} terpy complex [34], the use of terpy-derivatives for sensing of Fe^{2+} has mostly been performed in organic solvents or their mixtures with H₂O [11, 34-36], To the best of our knowledge, a limited amount of truly water-operational terpy-based metal sensing systems have been reported [19]. In this paper we report the design of a novel material based on terpy-functionalized screen-printed TiO₂ films, able to detect Fe^{2+} in water.

2. Experimental

2.1. Materials

Titanium (IV) isopropoxide (TIP) (Vertec, 97+%) and Ethylenediaminetetraacetic acid disodium salt dihydrate, 99+%, has been purchased from Alfa Aesar. Ethanol (absolute, \geq 99.8%), anhydrous terpineol, 5-15 mPa·s ethyl cellulose (48.0-49.5% w/w ethoxyl basis), 30-70 mPa·s ethyl cellulose (48.0-49.5% w/w ethoxyl basis), 30-70 mPa·s ethyl cellulose (48.0-49.5% w/w ethoxyl basis), titanium(IV) oxide, anatase nanopowder, <25 nm particle size were purchased from Sigma Aldrich. Aeroxide VP P90 fumed titanium dioxide was kindly donated by Evonik. 2,2':6',2"-terpyridin-4'-ylphosphonic acid was synthesized according previously published procedures [25, 37-39].

2.2. Methods

2.2.1. The morphology of SAM-anchored surfaces

STM images were recorded on Nanosurf NaioSTM using Nanosurf Naio 3.4.0 software. All STM images were obtained in the constant current mode by applying a tunneling current I_t of 70 to 250 pA and a sample bias V_t of 200 to 800 mV. The STM tips were mechanically cut from Pt/Ir wire (80/20, diameter 0.25 mm, Nanoscience). The calibration of the piezoelectric positioners was verified by atomic resolution imaging of graphite.

2.2.2. Surface coverage determination

UV-vis spectroscopy was used to determine the molecular density of monolayer films deposited on 1 mm quartz substrates as previously reported [40]. The instrument used for these experiments was a dual-beam Varian Cary® 50 UV-Vis spectrophotometer, with a modified sample holder for thin film measurements. Assuming uniform distribution of FeL₂ molecules on the surface and expecting the extinction coefficient in solution and in a monolayer is the same, we can relate the absorbance spectra with a calculated surface concentration, rewriting the Beer–Lambert law as follows:

$$\Gamma_{monolayer}[molecules \cdot cm^{-2}] = \frac{A \cdot 10^{-3} [cm^{-3}] \cdot N_{Av}[molecules \cdot mol^{-1}]}{2 \cdot \varepsilon [M^{-1} cm^{-1}]};$$
(1)

Here $\Gamma_{monolayer}$ is surface concentration, *A* is monolayer absorbance, N_{Av} is Avogadro's number, and ε is molar extinction coefficient. The factor of 2 in denominator was implemented since quarts slides were functionalized from both sides.

The peak at 560 nm characteristic to $\text{Fe}\mathbf{L}_2$ gives a molar extinction coefficient in solution of 11970 M⁻¹ cm⁻¹. The monolayer absorbance for this FeL₂ sample was 0.0029.

 $\Gamma_{monolayer} = \frac{0.0029 \cdot 10^{-3} [cm^{-3}] \cdot 6.02 \cdot 10^{23} [molecules \cdot mol^{-1}]}{2 \cdot 11970 [M^{-1} cm^{-1}]} = 7.29 \cdot 10^{13} [molecules \cdot cm^{-2}] \approx 0.7 [molecules \cdot nm^{-2}]$ This corresponds to area of 1/0.7=1.43 nm² per molecule. That is close to 1.21 nm² per molecule determined from the crystal structure [41].

The peak at 313 nm was chosen to estimate surface coverage of the ligand. This peak of \mathbf{L} gives a molar extinction coefficient in solution of 4910 M⁻¹ cm⁻¹. The monolayer absorbance for this \mathbf{L}

sample was 0.002. Analogously, surface concentration of the L is 1.23 [molecules nm^{-2}] that corresponds to $1/1.23=0.81 nm^2$ per molecule.

2.2.3. Anchoring of L onto TiO₂ Nanoparticles: Preparation of L-TiO₂ NPs

Immobilization of **L** was accomplished by stirring of the suspension of TiO_2 anatase nanoparticles (200 mg) with the solution of **L** (40 mg, 0.127 mmol) in DI water (40 mL). The mixture was stirred for 24 hours at room temperature. After, the suspension was separated by centrifugation and the liquid part was removed. The solid residue was thoroughly washed with DI water (3 x 50 mL), centrifuged and decanted, washed with ethanol (3 x 50 mL), and dried in vacuum for 24 hours.

2.2.4. Reaction of L-TiO₂ NP with Fe²⁺ solution.

Suspension of L-TiO₂ NPs was reacted with 40 mL of 45 ppm solution of Fe²⁺ in DI water. The mixture was stirred for 24 hours at room temperature. TiO₂ nanoparticles change colour from white to deep pink within 30 sec. Further stirring in the solution of Fe²⁺ in DI water had no influence on the NP colour. After, the mixture was separated by centrifugation. Deep magenta solid residue was thoroughly washed with DI water (6 x 50 mL), and ethanol (3 x 50 mL), centrifuged and decanted. Then solid precipitate was dried in vacuum for 24 hours.

2.2.5. Characterization of bare and L-TiO₂ anatase NPs

Bare and L-TiO₂ anatase NPs were characterized using Hitachi H7000 Transmission Electron Microscope operating under iTEM 5.2 software.

2.2.6. Syntheses and deposit of titanium dioxide scaffolds

Mesoporous titanium dioxide beads (TiO₂ NPs) have been obtained by hydrothermal synthesis according to the procedure reported in Chen et al. [42] and Latini et al. [43]. Briefly, titanium tetraisopropoxide hydrolysis has been conducted in hydroethanolic medium by using hexadecylamine and KCl as templating agent and ionic strength buffer, respectively. Autoclaving and annealing conditions have been previously reported [43]. Screen printing paste comprised of titanium dioxide mesoporous beads (scaffold material), ethyl cellulose (binder and thickener) and α -terpineol (solvent) has been prepared according to Ito et al. [44] and screen printed on clean soda lime glass or FTO-covered glass. Scaffold's film thickness was adjusted by one-by-one printing of several layers. Furthermore, commercially available high surface area P90 titanium dioxide was implemented into the paste as a scaffold material and screen printed for comparison.

2.2.7. Mesoporous TiO₂ NPs characterization

XRD.Powder X-ray diffraction analysis of mesoporous TiO₂ beads has been carried out by using a PanalyticalX'Pert PRO MPD diffractometer equipped with Cu K α source ($\lambda = 0.154184$ nm) and a X'Celerator 1D RTMS silicon strip detector. The angular resolution (in 2 θ) is 0.001°. Beam divergence, peaks asymmetry, and beam width have been adjusted by using a 0.04 rad soller slit, a 1° divergence slit and a 20 mm mask on the incident beam path. Low background level and beam

collimation have been assured by using a 6.6 mm anti-scatter slit and a 0.04 rad collimator on the diffracted beam path. Rietveld refinement analysis has been performed through MAUD software [45]. SEM.-Scanning electron microscope images of the samples have been collected by using a Zeiss Auriga FE-SEM microscope.Surface area and porosimetry. Specific surface area has been determined by using an ASAP 2020 Accelerated Surface Area and Porosimetry Analyzer (Micrometrics Instrument Corporation, U.S.) through the BET (Branauer-Emmett-Teller) procedure. Pore size distribution has been in the mesoporous range has been determined by the BJH (Barret-Joyner-Halenda) method by the desorption branch of the isotherm.

2.2.8. Anchoring of L onto TiO2 screen-printed films

Screen-printed films of TiO₂ on glass were immersed into solution of L (20 mg, 0.064 mmol) in 100 mL of DI water. After 30 min films were copiously washed with DI water and isopropanol (4 x 4 mL) and dried by stream of air or N₂. The same procedure was applied after each regeneration cycle.

2.2.9. Characterization of screen-printed films

Profilometry.–Tencor Alpha-Step-200 profilometer was equipped with a standard stylus of 12.5 micron radius. Sample leveling was automatically computed after each scan and digitized using GetData Graph Digitizer 2.24 software. Typical scan lengths were 10,000 um.Uv-Vis Diffuse Reflectance Spectroscopy. Diffuse reflectance measurements have been performed on the bare, *N*-aromatic ligand sensitized and Iron adsorbed films. The spectra have been collected in the range 280-1200 nm in transmittance mode using a double-beam Lambda 750S spectrophotometer (Perkin-Elmer, U.S.) equipped with a 60 mm integrating sphere. X-Ray Photoelectron Spectroscopy.XPS measurements were performed using a Thermo Scientific K-Alpha surface analysis system equipped with a monochromated Al Ka X-ray source (1486.7 eV). Ultra high vacuum (UHV) conditions (10⁻⁹ to 10^{-10} Torr) were maintained during the experiment. The instrument is fitted with a flood source for an effective charge compensation, The Ti⁴⁺ 2p3/2 peak of TiO₂ at 458.6 eV was used as an internal reference peak for calibrating the binding energy. A Smart fit algorithm was used for background subtraction and a Powell peak-fitting algorithm for data analysis. The elemental ratio was determined using Wagner atomic sensitivity factors [46].

3. Results and discussion

It is well documented that the terpy core motif allows visual differentiation of iron cations due to a rapid color change from colorless to deep magenta and this effect has been widely employed for the determination of Fe²⁺. Remarkably, after more than 80 years since the discovery of the first Fe²⁺ terpy complex [34], the use of terpy-derivatives for sensing of Fe²⁺ has mostly been performed in organic solvents or their mixtures with H₂O [11, 34-36], To the best of our knowledge, a limited amount of truly water-operational terpy-based metal sensing systems have been reported [19]. Recently, we utilized 2,2':6',2"-terpyridin-4'-ylphosphonic acid (L), which is able to interact with the number of metal ions via its *N*-aromatic terpy moiety, for the selective detection of Fe²⁺, Fe³⁺, Ru³⁺ and Zn²⁺ in aqueous media [41]. Reliable and fast quantification of 50 ppb to 1 ppm levels of each target cation was demonstrated in solution. In this paper the present communication we report the embedding of terpy-based water soluble receptors onto hydrophilic glass, fluorine-doped tin oxide

(FTO) coated glass, TiO₂ nanoparticles, and screen-printed porous surfaces, and explore the performance of these novel materials as Fe^{2+} sensors. The phosphonate functional group not only makes the ligand water soluble, but allows for the formation of robust self-assembled monolayers on a variety of surfaces. Thus phosphonates have been utilized for the surface modification of a wide range of oxide substrates [47] such as titania [48, 49], alumina [50], iron oxides [51], hafnium oxide [52], indium-tin oxide (ITO) [53], and silica [54]. Phosphonate-anchored SAMs are easily prepared in air, and form well-ordered strongly bound films on oxide surfaces with molecular densities close to those found in single crystals [55]. Taking this into account, we prepared the complex FeL₂ and deposited it onto FTO/glass and quartz surfaces (Fig. 1). Scanning tunneling microscopy (STM) analysis shows that the roughness of the FeL₂ functionalized FTO/glass surface (Fig 1E-H) is very similar of that of unfunctionalized FTO/glass (Fig 1 A-D), which suggest the formation of a monolayer. The embedding of the complex on the glass, quartz or FTO/glass surfaces can be easily observed by the naked eye. On the contrary, ligand anchoring of L on quartz surfaces does not result in any visible colour. However, this process could be monitored by UV-vis spectroscopy which confirms the formation of a monolayer of L on the surface (Fig. 1J). Moreover, insertion of quartz slides with the pre-deposited L ligand in 0.1 mM solution of Fe²⁺ results in insignificant changes in UV-vis spectra of the quartz substrates. Further immersion of these slides into a 0.1 mM solution of the ligand does not change the UV-vis spectra of surface-bonded layer at 560 nm, which is characteristic of a metal-to ligand charge transfer (MLCT) transition for a FeL₂ type complex [56], suggesting that Fe^{2+} did not incorporate into the ligand anchored to the surface. In contrast, preparation of the FeL_2 complex in solution followed by its deposition on quartz slides results in a significant increase of intensities at 287 nm and 332 nm, as well as the appearance of a new peak at 569 nm (see Fig. 1J). The red shift of the on-surface FeL_2 monolayer with respect to FeL_2 in aqueous solution (278, 321 and 560 nm, respectively) is the consequence of the change in the microenvironment of FeL_2 in these two different media [57].

Further characterization of the FeL₂ complex deposited on glass surface by XPS (Fig. 1I) verifies the presence of iron, nitrogen, carbon and phosphorous on the surface within ratios close to the molecular stoichiometry. The binding energy of any XPS peak depends on the oxidation state of an element and on its binding site on the surface. Thus the binding energy of the Fe $2p_{3/2}$ peak is located at 709.6 eV; the binding energy of Fe $2p_{1/2}$ can be seen at 722.1 eV; both peaks are characteristic for Fe²⁺. The N 1s peak at 400.9 eV is evidently associated with aromatic nitrogen that provides the electron density to form the complex. The C 1s peak is centered at 286.0 eV, which is in a good agreement with aromatic carbon. An asymmetric broad peak observed at 133.6 eV is due to the overlapping of the P $2p_{3/2}$ and P $2p_{1/2}$ components, which is characteristic for phosphates.

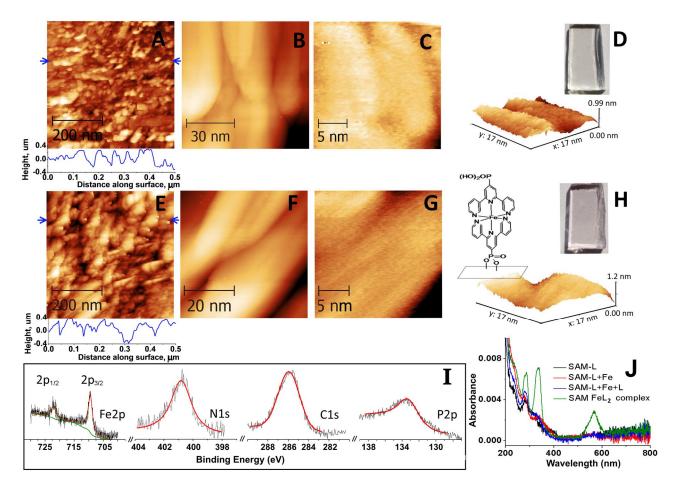


Figure 1. Representative STM images: (**A-D**) bare FTO/glass and (**E-H**) SAM-FeL₂ on FTO/glass slides. Representative XPS data for SAM-FeL₂ on glass slides (**I**): Fe(2p), N(1s), C(1s), and P(2p). The black line shows the experimental data, while the colored lines are the overall fitted spectra. UV-vis spectra (**J**) of as-deposited SAM-L (black), followed by its step-by-step treatment with Fe²⁺ aqueous solution (red) and solution of L (blue) show no significant change of the spectra suggesting no FeL₂ complex formation. In comparison, UV-vis spectrum of FeL₂ complex pre-formed in solution and then deposited as a SAM on quartz surface (green) show a distinguishable characteristic peak at 569 nm.

To estimate the packing density of FeL₂ molecules on glass surfaces, the two-dimensional surface concentration $\Gamma_{\text{monolayer}}$ (molecules cm⁻²) was calculated based on UV-vis data [40]. For this FeL₂ solution molar extinction coefficient of $\varepsilon_{560\text{nm}}$ = 11 970 M⁻¹ cm⁻¹ and the absorbance observed for the monolayer (see Fig. 1J and experimental section) resulted in a $\Gamma_{\text{monolayer}}$ of 7·10¹³ molecules/cm². This corresponds to a surface area occupied by one molecule of FeL₂ equal to 1.43 nm². The surface area estimated from the crystal structure of FeL₂ is close to 1.2 nm² per molecule. The surface concentration of **L** was found to be 1.23 molecules/nm² that corresponds to 0.81 nm² per molecule. This suggests that the ligand forms well-ordered and dense packed monolayers on relatively smooth surfaces and thus due to the intermolecular π - π interactions the rotation of the terpy moieties that are anchored to the surface is limited. Reduced flexibility of the terpy moieties on the surface, in turn, restricts the participation of two neighboring surface-anchored ligands in complex formation.

We proposed that when a high specific surface area substrate is functionalized with the ligand, a certain fraction of the ligand molecules might be available for FeL₂ complex formation. Indeed, modification of commercially available titanium (IV) oxide anatase nanopowder by L followed by treatment of the functionalized L-TiO₂ NPs with Fe²⁺ solutions results in significant colour change (Fig. 2, A&B insets). Particle size distribution of the nanopowder functionalized by L before (Fig. 2A) and after treatment with Fe²⁺ (Fig. 2B) remains very similar and close to 25 nm in diameter as declared for bare TiO₂ nanopowder by the manufacturer.

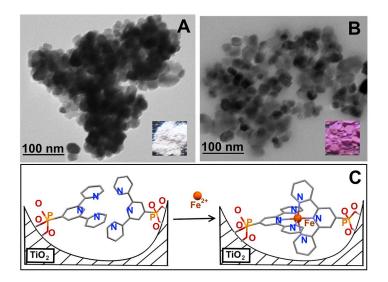


Figure 2. Representative TEM images of commercial TiO₂, anatase nanopowder, functionalized by **L**. **A**: before and **B**: after treatment with Fe^{2+} aqueous solutions. **C**: Proposed reaction mechanism of Fe^{2+} interactions with **L** molecules bound to TiO₂ nanopowder surface.

To make this system more practical and applicable for easy and rapid Fe²⁺ detection, a paste comprised of high (70-110 m^2/g) specific surface area Evonik P-90 commercial TiO₂ nanopowder (scaffold material), ethyl cellulose (binder and thickening agent) and α -terpineol (solvent) has been prepared according to previously reported procedure [44]. The paste was screen printed on base piranha solution pre-treated soda lime glass as 1x1 cm squares, dried and thermally treated in air. One, three and six layers (2 µm each) were screen printed one-by-one to increase the volume of the detecting cell and thus tune the detection range. When prepared, layered screen printed P-90-TiO₂ films were functionalized via immersion into a 0.65 mM aqueous solution of L for one hour followed by washing with isopropyl alcohol (3x3 mL) and dried by a stream of air. The resulting sensing strips were treated with 150 µL of 45 ppm Fe²⁺ aqueous solution; all sensing strips turned deep magenta with comparable intensity. These results confirm that novel material can detect Fe²⁺ in an aqueous solution. However, we believe that P90 small particles size (typically 15 nm) favors sintering processes resulting in the formation of surfaces where diffusion through the layers is impeded. As such only a reduced amount of surface remains accessible for the functionalization by the molecular receptor L, which is active in metal-ion recognition. As a result, P90-TiO₂ based screen-printed materials do not allow reasonable control of sensitivity since no significant difference in colour intensity was observed between one and multiple (up to 6 layers) screen-printed films.

To validate our initial assumption regarding the role of nanoparticle dimensions and pore size, we synthesized TiO₂ nanoparticles (crystallites size estimated by XRD ~ 25 nm) arranged in large (1263±86 nm in diameter) mesoporous (Fig. 3A) hierarchical spheres (beads). The presence of mesopores (IUPAC classification 2-50 nm) is easily recognizable by the characteristic hysteresis in the N₂ adsorption-desorption graph (Fig. 3B). Specific surface area determined by the BET (Branauer-Emmett-Teller) method has been found to be 78.4 ± 0.4 m²/g. Pore size distribution was determined by the BJH (Barret-Joyner-Halenda) method (Fig. 3D) and it has revealed a pore diameter distribution peaking at ca. 18 nm. Thus, although specific surface area of synthesized mesoporous TiO₂ nanoparticles is comparable to the P-90 one, the former are characterized by an almost monodisperse porosity stemming from spherical organization of randomly oriented nanocrystals. Moreover, the spherical shape of mesoporous TiO₂ beads minimize the surface energy tend to avoid surface shrinking during the annealing process. Further analysis of the synthesized mesoporous TiO₂ material by X-ray powder diffraction followed by Rietveld refinement analysis software)[45] results in the following lattice parameters: $a = 3.782 \pm 0.001$ Å; (MAUD $c = 9.492 \pm 0.003$ Å. The data are in good agreement with the reference data (a = 3.784479 Å, c = 9.5137 Å) for the tetragonal crystal structure of anatase (space group i41/amd).

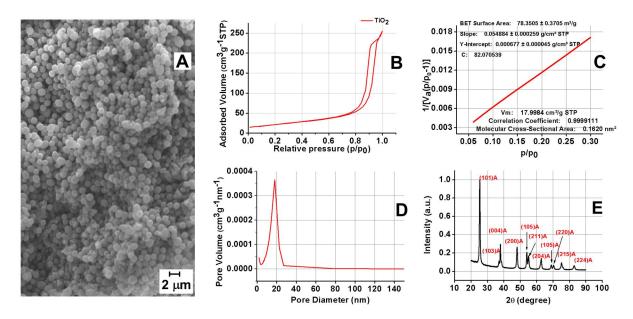


Figure 3. A: Typical SEM image of synthesized mesoporous TiO_2 NPs. B: Adsorption-desorption isotherm for mesoporous TiO_2 NPs. C: BET linear region fit for TiO_2 mesoporous beads. D: Pore diameters distribution obtained by BJH method. E: Powder diffraction spectrum of mesoporous TiO_2 beads.

Synthesis of mesoporous beads followed by their layer-by-layer additive screen-printing and functionalization by **L**, results in Fe²⁺ detecting films with a well-tunable thickness (Fig. 4). These mesoporous screen printed materials, in contrast to ones made from commercially available P90 TiO₂ NPs, benefit from better water diffusion and could be applied for quantitative Fe²⁺ detection. Dropwise addition of 150 μ L of a 45 ppm aqueous solution of Fe²⁺ on 1, 2, and 3 layered screen printed films of mesoporous TiO₂ nanopowder results in a rapid colour change from white to 3

shades of purple starting from less intense for the 1 layered film to most intense for the 3 layered one (Fig. 4). We think both mesoporosity and particle size are important factors allowing for modulation of sensitivity and better quantitative analysis.

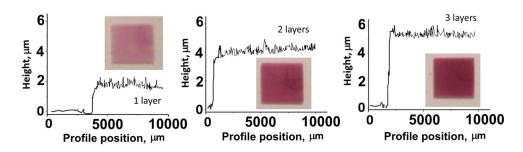


Figure 4. Typical profilometry curves of the mesoporous TiO_2 nanopowder deposited and sintered on FTO glass substrates show thickness of each layer of 2 um. Inserts: the images of corresponding films after applying of 150 µL of 45 ppm Fe²⁺ aqueous solution.

Three layered (6 um height) screen-printed films of mesoporous TiO_2 functionalized by **L** (further – detecting films) were further studied to determine the detectable Fe^{2+} concentration range. Applying 150 µL of 0.2 ppm, 2 ppm, and 45 ppm solutions of Fe^{2+} to the detecting films results in the appearance of a peak at 560 nm on the diffuse reflectance UV-Vis spectra (Fig. 5). Slight colour change takes place in the case of 0.2 ppm Fe^{2+} solution, but a naked eye distinguishable colour change was observed when 2 ppm and 45 ppm solutions have been used.

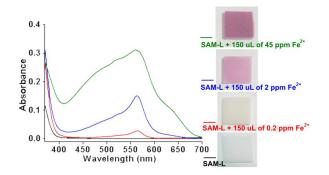


Figure 5. Diffuse reflectance UV-Vis spectra and optical photographs of mesoporous TiO₂ screen printed film (3 screen printed layers) functionalized by **L** and after treatment with 150 μ L of Fe²⁺ aqueous solutions. The characteristic peak at 560 nm confirms the formation of the FeL₂ complex.

It should be mentioned that according to the U.S. Environmental Protection Agency secondary drinking water standard and Guidelines for Canadian drinking water quality, maximum contaminant level of iron is 0.3 ppm; the United Nations Food and Agriculture Organization recommended level for irrigation waters is 5 ppm. On the other hand, iron in groundwater (in ferrous i.e. Fe^{2+} form) vary from 0.1 to 10 ppm [58]. Needless to say, the development of inexpensive, reusable and reliable materials which are able to rapidly detect iron in field conditions is in high demand. To insure our detecting films can be used as a material for Fe^{2+} determination, certain amounts of Fe^{2+} aqueous solutions were added to each film creating an appropriate coloring wheel (Fig. 6).

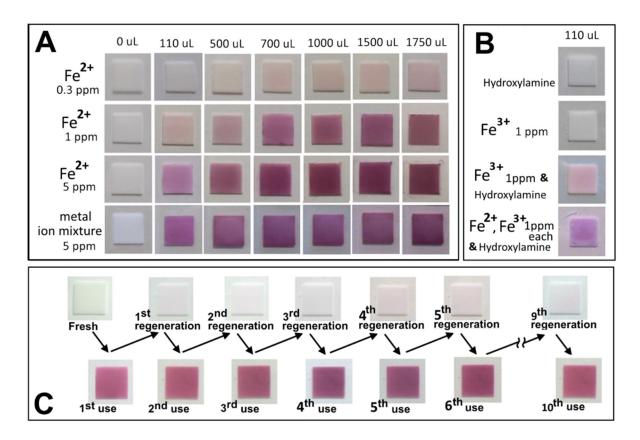


Figure 6. A: Colour wheel showing the colour change of detecting film after applying certain volumes of Fe^{2+} aqueous solutions: 0.3 ppm (upper row), 1ppm (2nd line from the top), 5 ppm (3rd line from the top), and a mixture of metal ions (including Fe^{2+}) 5 ppm each (bottom row) see main text for details. **B:** Water solutions of hydroxylamine do not change the colour of the detecting film, but reduce Fe^{3+} to Fe^{2+} making possible the detection of total iron in solution. **C:** Detecting films can be recovered by EDTA and reused, see main text for details.

Colour wheel results are consistent with our expectations: the colour intensity depends on the amount of Fe^{2+} loaded. This permits the visual quantification of iron cations in aqueous solutions ranging from 0.3 ppm to 5 ppm. To investigate the detecting films' selectivity towards Fe^{2+} in the presence of potentially interfering ions, an aqueous solution containing the following ions: Na⁺, Fe^{2+} , Cr^{3+} , Sn^{2+} , Ca^{2+} , Li^+ , Co^{2+} , Mg^{2+} , Cu^{2+} , Cs^+ , Ni^{2+} , Ba^{2+} , K^+ , and Mn^{2+} (5 ppm each) was prepared and portionwise applied to the detecting films. Fig. 6A clearly shows similar colour intensity of the detecting films after dripping certain volumes of 5 ppm solutions containing solely Fe^{2+} and Fe^{2+} in a mixture with thirteen other metal ions. This shows the material is highly selective to Fe^{2+} determination.

In addition, we studied the ability of our screen-printed material to detect Fe^{3+} ions in the aqueous solutions. Despite of the fact that aqueous solutions of Fe^{3+} do not change colour of the detecting film, Fe^{3+} can be detected in aqueous solutions if reduced to Fe^{2+} by hydroxylamine [59] (Fig. 6B).

Notably, the detecting films can be reused by immersing for 48 hours into a 0.5 mM solution of ethylenediaminetetraacetic acid disodium salt (EDTA). Iron release from the films into the EDTA

solution can be monitored by the visual color change of the films. Figure 6C shows color films behavior for 10 successful cycles of resetting, no evidence of sensitivity degradation can be detected. Increasing the temperature of the EDTA solution to 80°C reduces the film recovery time to 24 h.

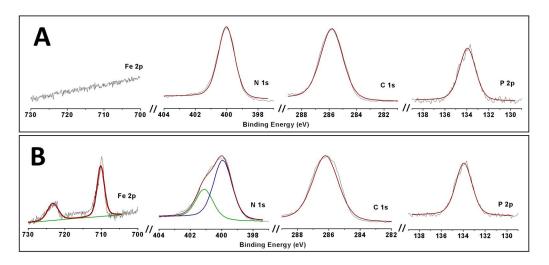


Figure 7. Comparative X-ray photoelectron spectra of the L anchored to mesoporous TiO₂ screen printed films A: before and B: after applying of 110 μ L of 5 ppm Fe²⁺ aqueous solution.

XPS analysis of detecting films before interaction with Fe²⁺ confirms the presence of N, C, and P of L on the surface (Fig. 7A). The sharp (with full width at half-maximum (FWHM) of 1.9 eV) N 1s peak is centered at 400.0 eV. The C 1s peak is centered at 285.8 eV, as expected for aromatic carbon. An asymmetric broad peak observed at 133.8 eV is characteristic for P 2p. The N:C:P ratio of 2.5:17.4:1.0 is close to 3:15:1 of **L** molecular stoichiometry. When 110 μ L of 5 ppm Fe²⁺ aqueous solution was applied to the detecting film (Fig 7B) the appearance of two new peaks was observed. The peaks with the binding energies of 709.9 eV and 722.4 eV are characteristic for Fe 2p_{3/2} and of Fe $2p_{1/2}$ of Fe²⁺, respectively. Interestingly, the P 2p peak position remains within the experimental error the same (133.8 eV) as before Fe^{2+} solution application, the C 1s peak is insignificantly shifted to 286.1 eV, but N 1s peak undergoes a significant change and now can be deconvoluted using a Powell peak-fitting algorithm with full width at half-maximum (FWHM) of 1.3 eV yielding a peak centered at 400.0 eV and a new peak centered at 401.1 eV. This value is close to 400.9 eV, which was observed for the FeL₂ complex deposited on the FTO surface (Fig. 1I). The P:Fe ratio is 1:0.01suggesting that as expected, just a part of L molecules is able to form FeL_2 complex. On the other hand, assuming that the complex formation takes place not on the flat surface, but inside the pores, the iron content can be underestimated due to the signal attenuation.

4. Conclusions

We observed that 2,2':6',2"-terpyridin-4'-ylphosphonic acid, **L**, forms dense SAMs on FTO/glass and quartz surfaces. Due to the close packing of the **L** molecules, these SAMs, once formed, cannot react with Fe^{2+} to form the FeL₂ complex. On the other hand, in-solution, pre-formed the FeL₂ complex forms dense SAMs on glass surface which are observable by the naked eye as pink colouration of the glass. Depositing of SAMs of **L** on high surface area TiO₂ nanostructures results

in a situation when some of the Ls are able to form FeL_2 complex. The formation of the Fe^{2+} complex in turn leads to significant colouration of the material. To improve detection abilities and adjust the Fe^{2+} detection limits, large mesoporous TiO_2 beads have been synthesized and screenprinted on glass slides. Functionalization of these beads by L results in reusable detecting films able to quantify Fe^{2+} in aqueous solutions with the concentration range from 0.3 ppm to 5 ppm. The selectivity of Fe^{2+} detection in the presence of 13 other metal ions was shown. This material is expected to be useful for the design of more advanced efficient optical, chemical, and biological sensors. Such sensors would benefit from high accuracy, reliability, portability and cost effectiveness.

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